



Figure 1: *Physalis angulata* (A) asymptomatic and (B) exhibiting typical begomovirus symptoms (apical chlorosis).

245x143mm (150 x 150 DPI)

1       **Identification of *Physalis angulata* (Solanaceae) as a Natural Alternative Weed**  
2                               **Host of Tomato Severe Rugose Virus in Brazil**

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10       A diverse array of *Begomovirus* species (family *Geminiviridae*) can induce economically  
11       important diseases in all major tomato-producing areas in Brazil (Fernandes et al., 2008).  
12       Weeds of the Solanaceae family have been identified as important alternative hosts of the  
13       viral species reported infecting tomato crops in the country (Barreto et al., 2013). In 2016,  
14       about 10% of the plants of the weed *Physalis angulata* L. were observed with  
15       begomovirus-like symptoms (apical chlorosis and stunting; Supplementary Fig. 1) around  
16       and within tomato fields in Venda Nova do Imigrante–ES (Southeast Brazil). Leaf tissue  
17       was collected from ten symptomatic and five healthy-looking plants, respectively. Total  
18       DNA was extracted using the CTAB method in combination with organic solvents  
19       (Boiteux et al., 1999). PCR assays with the universal begomovirus primers for DNA-A  
20       and DNA-B detection (Rojas et al., 1993) were carried out using total genomic DNA  
21       samples (20 ng/μL) as templates. Amplicons of ≈ 1,100 bp and ≈ 550 bp were obtained  
22       only in the ten symptomatic samples, which are in agreement with the expected fragment  
23       sizes of DNA-A and DNA-B from begomoviruses (Rojas et al. 1993). These amplicons  
24       were directly Sanger sequenced at the Embrapa Vegetables Genomic Analysis  
25       Laboratory. BLASTn analyses of the amplicon sequences indicated high nucleotide  
26       identities (99%, 98%) with DNA-A and DNA-B of tomato severe rugose virus (ToSRV)  
27       isolates (accession numbers JX415196, MG837739) from Brazil, suggesting the infection  
28       of ToSRV in these plants. To fully prove the infection and to characterize the ToSRV  
29       isolate associated with *P. angulata*, the DNA samples were submitted to rolling circle  
30       amplification (RCA) method followed by Nanopore sequencing (Oxford Nanopore  
31       Technologies), as previously described (Naito et al., 2019). The full-length DNA-A  
32       component of one isolate obtained from *P. angulata* (MN059848) was 98% identical to  
33       the DNA-A genome of a Brazilian ToSRV isolate from tomato (JX415196). The  
34       complete DNA-B sequence from the *P. angulata* isolate (MN059849) was 96% identical

35 to the DNA-B genome of a tomato-infecting ToSRV isolate obtained in Southeast Brazil  
36 (MG837739). The complete DNA-A component (2,592 nts) displayed all genomic  
37 features of the New World species, with one virion-sense ORF (AV1) and four  
38 complementary sense ORFs (AC1, AC2, AC3, and AC4). The complete DNA-B  
39 component (2,571 nts) displayed two ORFs, virion-sense (BV1) and complementary  
40 sense (BC1). The common regions of DNA-A and DNA-B components were identified  
41 (183 nts in length) and the presence of iterons was confirmed. ToSRV is so far the most  
42 prevalent tomato-infecting begomovirus in Brazil (Fernandes et al., 2008). To our  
43 knowledge, this is the first report of natural infection of *P. angulata* by ToSRV,  
44 expanding the viral host range to this widespread tropical and subtropical weed.  
45 Considering that *P. angulata* may function as a relevant natural source of ToSRV  
46 inoculum to tomato crops across tropical and subtropical regions, control of the weed  
47 should be a part of the integrated management of ToSRV in tomato crops.

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#### 49 **References**

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