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- 2 First report of *Dematophora bunodes* causing root rot of taro (*Colocasia esculenta*)
- 3 and leatherleaf fern (Rumohra adiantiformis) in Brazil
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- 11 Colocasia esculenta, taro (T), is a major staple food crop in the tropics, including Brazil.

 12 Provedera adjustiferania leatherhead form (LE) is breadly sultivated for its arramental
- 12 Rumohra adiantiformis, leatherhead fern (LF), is broadly cultivated for its ornamental
- fronds that are used as a component of flower arrangements. Soft root rot of T and LF,
- and accompanying rapid plant wilt and death, was observed in plantations in Espírito
- 15 Santo (Brazil), at Venda Nova do Imigrante, in April 2014 (LF) and July 2015 (T). Great
- losses were observed. Firstly, a few individual scattered plants showed symptoms of
- 15 losses were observed. Thistry, a few marviadar seattered plants showed symptoms of
- disease in the plantations, then aggregates of plants and, after a few seasons, the majority
- of the plants in the field died before harvest, leading to the abandonment of the activity
- by farmers. A white mycelial matt was observed on the crown and roots of dying T and
- 20 LF plants. Infected corms become necrotic and dark brown mycelial strands were
- 21 observed internally in tissues. Diseased organs were carefully washed and surface
- sterilized in 10% sodium hypochlorite. Samples of tissue were removed from the
- boundary of necrotic tissues and placed on potato dextrose-agar (PDA) plates and
- incubated at 23±2 C in the dark. Homogeneous mycelial colonies were isolated from
- both T and LF and, upon observation of microscope mounts under an Olympus BX 53
- 26 light microscope, pear-shaped hyphal swellings at the septae (Castro et al. 2013) were
- 20 light interoscope, pear-shaped hyphar swernings at the septac (Castro et al. 2013) were
- observed. A representative isolate from each host was deposited in the local culture
- collection as COAD 2911 (LF isolate) and COAD 2912 (T isolate). Additionally, DNA
- 29 was extracted from each culture using the Wizard Genomic DNA Purification Kit
- 30 (Promega) and the internal transcriptional spacer region was PCR amplified using the
- 31 primers ITS5 and ITS1 (White et al. 1990). The amplicons were sequenced by
- 32 MACROGEN (http://www.macrogen.com). Consensus sequences were deposited in
- 33 GenBank: MW561595 (LF), MW561596 (T). Consensus regions were compared against
- other sequences available in Genbank. A BLASTn analysis resulted in LF and T
- sequences respectively 99% (526/531bp) and 98% (412/420 bp) identity with that of
- 36 Dematophora bunodes (MN984619). Additionally, a phylogenetic analysis of a selected
- sequence alignment was performed on the CIPRES webportal (Miller et al., 2010) using
- 38 MrBayes v.3.1.1 (Ronquist & Huelsenbeck, 2003). A phylogenetic tree was generated
- showing that the placement of LF and T isolates is in *D. bunodes* (Wittstein et al. 2020).

- 1 Pathogenicity tests were performed for LF and T isolates against their original hosts. For
- 2 inoculum, bags of twice-autoclaved parboiled rice were seeded separately with each
- 3 isolate, which were allowed to colonize the rice for two weeks. Four healthy young LF
- 4 and T plants were utilized. Two extra healthy plants grown in the same conditions, but
- 5 not inoculated, served as controls. Thirty g of *Dematophora*-colonized rice was placed
- 6 in direct contact with stems or roots of each LF or T plant. Plants were maintained in a
- 7 dew chamber for 48 h after inoculation and then transferred to a greenhouse bench. All
- 8 inoculated plants developed wilt and root rot and died after 15-20 days. Controls remained
- 9 healthy. White mycelial colonies were formed over tissues of diseased LF and T and
- upon observation under the microscope, typical pear-shaped swellings were observed in
- slides prepared from newly obtained pure cultures from LF and T. *Dematophora bunodes*
- 12 (formerly Rosellinia bunnodes) has a worldwide distribution and is well known as a
- polyphagous plant pathogen (Farr and Rossman, 2020) but has never been reported as a
- pathogen either of LF or T before in Brazil and worldwide. Its report on LF and T further
- expands an already large host-range and resolves the etiology of the disease on LF and T.

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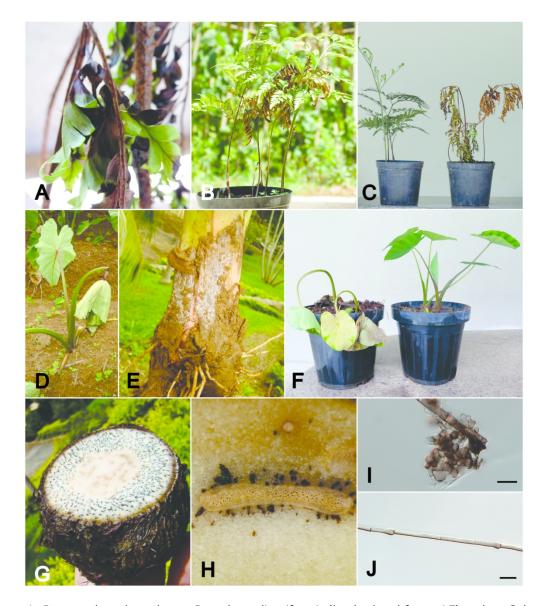


Figure 1. Dematophora bunodes on Rumohra adiantiformis (leatherhead fern – LF) and on Colocasia esculenta (taro – T). A-B. Foliage blight resulting from stem rot on LF. C. Potted LF showing results of Koch's postulates after fifteen days: left - healthy control vs. right - wilted inoculated plant. D. Wilted dying T plant in plantation. E. Collar rot on T uprooted individual. F. Potted T showing results of Koch's postulates after fifteen days: right - healthy control vs. left - wilted inoculated plant. G and H. Transversal section of diseased taro corm showing discoloration and soft root rot of central part and dark internal mycelial strands. I. Apresorial-pad of COAD 2911 (LF isolate) on PDA J. Hypha bearing typical Dematophora pyriform-swellings next to septae on COAD 2912 (T isolate).

209x237mm (300 x 300 DPI)

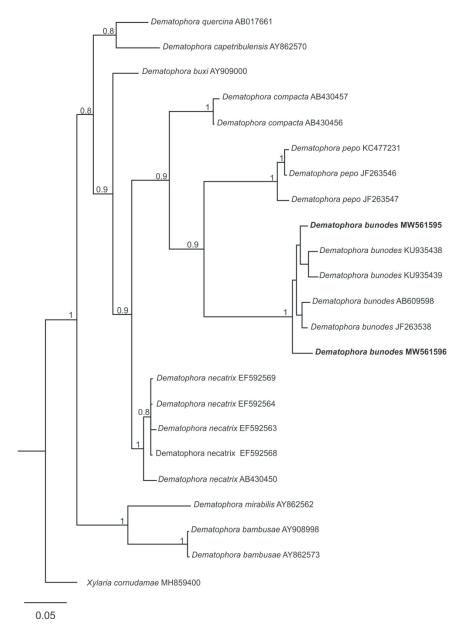


Figure 2. Phylogenetic tree generated by Bayesian analysis of ITS sequences showing the placement of COAD 2911 (leatherhead fern *Dematophora bunodes* isolate) and COAD 2912 (taro *D. bunodes* isolate) among species of *Dematophora*. Posterior probability values are indicated above the nodes. The isolates used in this study are highlighted in bold. The tree is rooted with *Xylaria cornudamae*.

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